PATENTS AND LITERATURE

ROBERT J. LINHARDT

Division of Medicinal and Natural Products Chemistry, College of Pharmacy, University of Iowa, Iowa City, IA 52242

The objective of this section is to keep readers aware of significant inventions and trends in industrial research as well as to highlight those areas of research that may lead to new biotechnological opportunities. In addition to DNA probes for clinical applications covered in the last issue, three other subject areas are being surveyed in 1986: protein engineering, mammalian cell culture, and microbial transformations. The subject of this, the fourth Patents and Literature Section of 1986 is protein engineering and site-directed mutagenesis.

Protein Engineering and Site-Directed Mutagenesis

Patents

This section identifies and gives a brief description of patents from the US patent literature from January 1975 through February 1986. The search headings were protein engineering, site-directed mutagenesis, and nucleotide modification or alteration. Both patent titles and abstracts were searched. Patent assignees were also searched for several of the major biotechnology companies. Copies of the US patents can be obtained for \$1.50 each from the Commissioner of Patents and Trademarks, Washington, DC 20231.

Bahl, C. P.

METHOD FOR SINGLE NUCLEOTIDE ALTERATION

US 4,351,901, Sep. 28, 1982

Assignee: Cetus Corp.

A method is described for altering a single nucleotide at a predetermined position in a gene involving the isolation of a single strand gene fragment extending up to the position before the nucleotide to be altered. A ribonucleotide or a protected deoxyribonucleotide corresponding to the desired altered nucleotide is attached at the end of this fragment. The fragment is then annealed to a complementary template that extends beyond the end of the fragment. The fragment is then extended complementary to the remainder of the template. The resulting partially mismatched double-stranded DNA is used to produce a pure DNA gene containing an altered deoxyribonucleotide at the single desired position.

Bender, R. and Duck, P. D.
CHEMICAL SYNTHESIS APPARATUS FOR PREPARATION OF
POLYNUCLEOTIDES

US 4,353,989, Oct. 12, 1982

Assignee: ens Bio Logicals Inc.

An apparatus is described for the stepwise synthesis of polynucleotides in which the polynucleotide chains are extended in stepwise fashion from a modified form of polymer support to which the first unit is linked. This apparatus includes a reaction column containing the polymer supported product, which acts as the reaction vessel, and a series of reaction bottles all connected by two-way valves arranged in series to the column by means of a fluid-flow conduit. The farthest upstream vessel of the series contains reaction solvent, used for washing purposes. The most downstream of the reaction vessels contain nucleotide reagents. Each of the values has two separate and discreet fluid-flow passageways, the first used exclusively for flow of reagent from the vessel and the second for flow of solvent or reagents from an upstream vessel. This eliminates the possibility of cross-contamination as a result of residual reagent left in the dead space of the valve. The valves are biased toward their solvent flow condition. Materials are drawn through the fluid flow conduit and values by suction.

Itakura, K.
RECOMBINANT DNA CLONING VEHICLE
US 4,356,270, Oct. 26, 1982

and

MAMMALIAN GENE FOR MICROBIAL EXPRESSION US 4,571,421, Feb. 18, 1986 *Assignee:* Genetech, Inc.

Recombinant microbial cloning vehicles comprising heterologous DNA coding for the expression of mammalian hormones and other polypeptides, including plasmids suited for the transformation of bacterial hosts, are described. The latter incorporate a regulon homologous to the host in its untransformed state, in reading phase with the structural gene for the heterologous DNA. Cloning vehicles are described that code for the microbial expression of various proteins, including (a) a polypeptide hapten and additional protein sufficient in size to confer immunogenicity on the product of expression, which may find use in raising antibodies to the hapten for assay use or in the manufacture of vaccines; and (b) a desired polypeptide product and additional protein from which the desired product may be cleaved. Methods of preparing synthetic structural genes coding for the expression of mammalian polypeptides in microbial cloning systems are also described.

Mark, D. F., Lin, L. S., and Lu, S. D. Y. HUMAN RECOMBINANT INTERLEUKIN-2 MUTEINS US 4,518,584, May 21, 1985
Assignee: Cetus Corp.

Muteins of biologically active proteins, such as IFN-Beta and IL-2, in which cysteine residues that are not essential to biological activity have been deleted or replaced with other amino acids to eliminate sites for intermolecular crosslinking or incorrect intramolecular disulfide bridge formation. These muteins are made by bacterial expression of mutant genes that encode the muteins that have been synthesized from the genes for the parent proteins by oligonucleotide-directed mutagenesis.

Paau, A., Platt, S. G., and Sequeira, L.
ASSAY METHOD AND PROBE FOR POLYNUCLEOTIDE
SEQUENCES

US 4,556,643, Dec. 3, 1985

Assignee: Agracetus

An assay method for the detection of a specific nucleotide target sequence in a polynucleotide test extract is disclosed that utilizes a polynucleotide-modified probe, including both a cDNA sequence substantially complementary to the specific target sequence and a protein binding sequence. The assay is conducted by exposing the modified probe to the polynucleotide test extract for hybridization and then exposing the complex to the protein that binds to the protein-binding sequence. An assay, such as an immunoassay, is then conducted on the test sample to indicate the presence of the specific target sequence by detecting the presence of the binding protein.

Paddock, G. V.

RECOMBINANT cDNA CONSTRUCTION METHOD AND HYBRID NUCLEOTIDES USEFUL IN CLONING

US 4,362,867, Dec. 7, 1982

US 4,503,151, Mar. 5, 1985

Assignee: Research Corp.

Compounds useful as complementary DNA (cDNA), including deoxyribonucleotides and at least one ribonucleotide, are described. They may be depicted by the general formula: 3'(rN) (b) (dN) (a) 5'5' (dN) (c) 3', where (dN) (a) and (dN) (c) represent series of deoxyribonucleotides and (rN) (b) represents a series of ribonucleotides; a, b, and c are the number of nucleotides in the series, and b is ≥ 1 , a is ≥ 35 , and c is ≥ 10 ; the series of deoxyribonucleotides (dN) (a) includes a series of deoxyribonucleotides that are substantially complementary to the series of deoxyribonucleotides (dN) (c). These compounds may be prepared from messenger RNA (mRNA) containing the genetic information necessary for cellular production of desired products, such as polypeptides. After appropriate modification, they may be combined with DNA from a suitable cloning vehicle, such as a plasmid, and the resulting combined DNA used to transform bacterial cells. The transformed bacterial cells may then be grown and harvested and the desired product or products recovered.

Riggs, A. D. METHOD FOR MICROBIAL POLYPEPTIDE EXPRESSION US 4,366,246, Dec. 28, 1982

and

MICROBIAL POLYPEPTIDE EXPRESSION VEHICLE US 4,425,437, Jan. 10, 1984

and

TRANSFORMANT BACTERIAL CULTURE CAPABLE OF EXPRESSING HETEROLOGOUS PROTEIN US 4,431,739, Feb. 14, 1984

and

METHOD AND MEANS FOR SOMATOSTATIN PROTEIN CONJUGATE EXPRESSION

US 4,563,424, Jan. 7, 1986 *Assignee:* Genetech, Inc.

Recombinant microbial-cloning vehicles comprising heterologous DNA coding for the expression of mammalian hormones and other polypeptides, including plasmids suited for the transformation of bacterial

hosts are described. The latter incorporate a regulon homologous to the

host in its untransformed state, in reading phase with the structural gene for the heterologous DNA. Cloning vehicles are described that code for the microbial expression of various proteins, including (a) a polypeptide hapten and additional protein sufficient in size to confer immunogenicity on the product of expression, which may find use in raising antibodies to the hapten for assay use or in the manufacture of vaccines; and (b) a desired polypeptide product and additional protein from which the desired product may be cleaved. Methods of preparing synthetic structural genes coding for the expression of mammalian polypeptides in microbial cloning systems are also described.

Rubin, H.
METHOD OF INSERTING AMINO ACID ANALOGS INTO PROTEINS
US 4,568,640, Feb. 04, 1986

A method is described for substituting one amino acid for another in a protein chain to improve selected properties of the protein. Initially, an mRNA molecule capable of producing an unmodified protein of the type it is desired to modify is obtained. Next, the codon along the mRNA chain that ordinarily accepts the anticodon corresponding to a first amino acid that is to be replaced with a selected second amino acid is determined. The tRNA that ordinarily brings the first amino acid to that site is modified to carry the second amino acid instead. The modified protein is then formed by translation techniques in the presence of the modified tRNA, whereby the second amino acid is incorporated in the protein chain in place of the first amino acid. Several methods of accomplishing the modification of the selected tRNA are also described.

Weissman, S. M., Pereira, D., and Sood, A. METHOD FOR CLONING GENES US 4,394,443, Jul. 19, 1983
Assignee: Yale University

A method is provided for isolating and identifying a recombinant clone having a DNA segment coding for at least one desired heterologous polypeptide, at least a short amino acid sequence of which is known. cDNA synthesis is performed on a mixture of mRNAs containing the mRNA coding for the desired polypeptide, the resultant cDNA mixture is isolated, inserted into recombinant cloning vehicles, the hosts are transformed with the vehicles, the transformants are separated, and a recombinant clone is isolated and identified that contains a DNA segment that is at least partially homologous to one oligonucleotide probe specific for the DNA segment. The probe is an extension of the nucleotide sequence of an oligonucleotide primer having a nucleotide sequence complementary to a region of the target mRNA coding for a portion of the known amino acid sequence and is complementary to a

longer region of the target mRNA coding for a longer portion of the known amino acid sequence. Recombinant clones coding for human histocompatibility antigens are described.

Literature

This section surveys literature in the area of protein engineering and sitedirected mutagenesis from January 1984 to April 1986. This section includes only selected articles that appeared during this time period.

REFERENCES

- 1. Asghar, S. S., and Ali, L. (1985), Possible Treatment of Some Genetic Deficiency Diseases—A Hypothesis. *Med. Hypotheses* **16**, 371–375.
- 2. Barany, F. (1985), Single-Stranded Hexameric Linkers: A System for In-Phase Insertion Mutagenesis and Protein Engineering. *Gene* 37, 111–123.
- 3. Carter, P., Bedouelle, H., and Winter, G. (1985), Improved Oligonucleotide Site-Directed Mutagenesis Using M13 Vectors, *Nucleic Acids Res.* **13**, 4431–4443.
- 4. Carter, P. J., Winter, G., Wilkinson, A. J., and Fersht, A. R. (1984), The Use of Double Mutants to Detect Structural Changes in the Active Site of the Tyrosyl-tRNA Synthetase (*Bacillus stearothermophilus*). *Cell* **38**, 835–840.
- 5. Chong, P., Hui, I., Loo, T., and Gillam, S. (1985), Structural Analysis of a New GC-Specific Insertion Element IS186. FEBS Lett. 192, 47–52.
- 6. Clune, A., Lee, K. S., and Ferenci, T., (1984), Affinity Engineering of Maltoporin: Variants with Enhanced Affinity for Particular Ligands. *Biochem. Biophys. Res. Commun.* **121**, 34–40.
- 7. Dalbadie-McFarland, G., Riggs, A. D., and Richards, J. H. (1984), Directed Mutagenesis as a Technique to Study Protein Function: Application to beta-Lactamase. *Biochem. Soc. Trans.* 12, 226–228.
- 8. Estelle, M., Hanks, J., McIntosh, L., and Somerville, C. (1985), Site-Specific Mutagenesis of Ribulose-1,5-Bisphosphate Carboxylase/Oxygenase. Evidence that Carbamate Formation at Lys 191 is Required for Catalytic Activity. *J. Biol. Chem.* **260**, 9523–9526.
- 9. Estell, D. A., Graycar, T. P., and Wells, J. A. (1985), Engineering an Enzyme by Site-Directed Mutagenesis to be Resistant to Chemical Oxidation. *J. Biol. Chem.* **260**, 6518–6521.
- 10. Fersht, A. R., and Winter, G. P. (1985), Redesigning Enzymes by Site-Directed Mutagenesis. *Ciba Found. Symp.* **111**, 204–218.
- 11. Fersht, A. R., Shi, J. P., Knill-Jones, J., Lowe, D. M., Wilkinson, A. J., Blow, D. M., Brick, P., Carter, P., Waye, M., and Winter, G. (1985), Hydrogen Bonding and Biological Specificity Analysed by Protein Engineering. *Nature* 314, 235–238.
- 12. Fersht, A. R., Shi, J. P., Wilkinson, A. J., Blow, D. M., Carter, P., Waye, M. M. Y., and Winter, G. P. (1984), Analysis of Enzyme Structure and Activity by Protein Engineering. *Angew, Chem. Ed. Engl.* **23**, 467–473.
- 13. Gething, M. J., Doms, R. W., York, D., and White, J. (1986), Studies on the Mechanism of Membrane Fusion: Site-Specific Mutagenesis of the Hemagglutinin of Influenza Virus. *J. Cell Biol.* 102, 11–23.

14. Gillam, S., Astell, C. R., Jahnke, P., Hutchison, C. A., and Smith, M. (1984), Construction and Properties of a Ribosome-Binding Site Mutation in Gene E of Luminal Diameter X174 Bacteriophage. *J. Virol.* **52**, 892–896.

- 15. Glickman, B. W., and Ripley, L. S. (1984), Structural Intermediates of Deletion Mutagenesis: A Role for Palindromic DNA. *Proc. Natl. Acad. Sci. USA* 81, 512–516.
- 16. Howell, E. E., Villafranca, J. E., Warren, M. S., Oatley, S. J., and Kraut, J. (1986), Functional Role of Aspartic Acid-27 in Dihydrofolate Reductase Revealed by Mutagenesis. *Science* **231**, 1123–1128.
- 17. Jay, E., MacKnight, D., Lutze-Wallace, C., Harrison, D., Wishart, P., Liu, W. Y., Asundi, V., Pomeroy-Cloney, L., Rommens, J., and Eglington, L., (1984). Chemical Synthesis of a Biologically Active Gene for Human Immune Interferon-gamma. Prospect for Site-Specific Mutagenesis and Structure-Function Studies. *J. Biol. Chem.* **259**, 6311–6317.
- 18. Kaiser, E. T., Lawrence, D. S., and Rokita, S. E. (1985), The Chemical Modification of Enzymatic Specificity. *Annu. Rev. Biochem.* **54**, 565–595.
- 19. Leatherbarrow, R. J., Fersht, A. R., and Winter, G. (1985), Transition-State Stabilization in the Mechanism of Tyrosyl-tRNA Synthetase Revealed by Protein Engineering. *Proc. Natl. Acad. Sci. USA* **82**, 7840–7844.
- 20. Liang, S. M., Thatcher, D. R., Liang, C. M., and Allet, B. (1986), Studies of Structure-Activity Relationships of Human Interleukin-2. *J. Biol. Chem.* **261**, 334–337.
- 21. Lowe, D. M., Fersht, A. R., Wilkinson J., Carter, P., and Winter, G. (1985), Probing Histidine-Substrate Interactions in Tyrosyl-tRNA Synthetase Using Asparagine and Glutamine replacements. *Biochemistry* **24**, 5106–5109.
- 22. Matthews, D. A., Bolin, J. T., Burridge, J. M., Filman, D. J., Volz, K. W., and Kraut, J. (1985), Dihydrofolate Reductase. The Stereochemistry of Inhibitor Selectivity. *J. Biol. Chem.* **260**, 392–399.
- 23. Matthews, D. A., Bolin, J. T., Burridge, J. M., Filman, D. J., Volz, K. W., Kaufman, B. T., Beddell, C. R., Champness, J. N., Stammers, D. K., and Kraut, J. (1985), Refined Crystal Structures of *Escherichia coli* and Chicken liver Dihydrofolate Reductase Containing Bound Trimethoprim. *J. Biol. Chem.* **260**, 381–391.
- 24. Maugh, T. H. (1984), Need a Catalyst? Design an Enzyme. *Science* **223**, 269–271.
- 25. Mead, D. A., Skorupa, E. S., and Kemper, B. (1985), Single-Stranded DNA SP6 Promoter Plasmids for Engineering Mutant RNAs and Proteins: Synthesis of a 'Stretched' Preproparathyroid Hormone. *Nucleic Acids Res.* **13**, 1103–1118.
- 26. Milstein, C. (1986), From Antibody Structure to Immunological Diversification of Immune Response. *Science* **231**, 1261–1268.
- 27. Nagai, K. (1985), Protein Engineering. *Tanpakushitsu Kakusan Koso* **30**, 950–962.
- 28. Newman, A. J., Lin, R. J., Cheng, S. C., and Abelson, J. (1985), Molecular Consequences of Specific Intron Mutations on yeast mRNA Splicing In Vivo and In Vitro. *Cell* **42**, 335–344.
- 29. Neuhaus, J. M., Soppa, J., Wright, J. K., Riede, I., Blocker, H., Frank, R., and Overath, P. (1985), Properties of a Mutant Lactose Carrier of *Escherichia coli* with a Cys148—Ser148 substitution. *FEBS Lett.* **185**, 83–88.
- 30. Nisbet, I. T., Beilharz, M. W., Hertzog, P. J., Tymms, M. J., and Linnane, A. W. (1985), Single Amino Acid Substitutions at Conserved Residues of Human Interferon-alpha can Effect Antiviral Specific Activity. *Biochem. Int.* 11, 301–309.

31. Nomura, A., Negishi, K., and Hayatsu, H. (1985), Direct-Acting Mutagenicity of N4-Aminocytidine Derivatives Bearing Alkyl Groups at the Hydrazino Nitrogens. *Nucleic Acids. Res.* **13**, 8893–8899.

- 32. Paluh, J. L., Zalkin, H., Betsch, D., and Weith, H. L. (1985), Study of Anthranilate Synthase Function by Replacement of Cysteine 84 Using Site-Directed Mutagenesis. *J. Biol. Chem.* **260**, 1889–1894.
- 33. Peden, K. W., and Pipase, J. M. (1985), Site-Directed Mutagenesis of the Simian Virus 40 Large T-Antigen Gene: Replication-Defective Amino Acid Substitution Mutants that Retain the Ability to Induce Morphological Transformation. J. Virol. 55, 1–9.
- 34. Petsko, G. A., Davenport, R. C., Jr., Frankel, D., and RaiBhandary, U. L. (1984), Probing the Catalytic Mechanism of Yeast Triose Phosphate Isomerase by Site-Specific Mutagenesis. *Biochem. Soc. Trans.* 12, 229–232.
- 35. Pielak, G. J., Mauk, A. G., and Smith, M. (1985), Site-Directed Mutagenesis of Cytochrome *c* Shows that an Invariant Phe is not Essential For Function. *Nature* **313**, 152–154.
- 36. Richardson, T. (1985), Chemical Modifications and Genetic Engineering of Food Proteins. J. Dairy Sci. 68, 2753–2762.
- 37. Robey, E. A., and Schachman, H. K. (1984), Site-Specific Mutagenesis of Aspartate Transcarbamoylase. Replacement of Tyrosine 165 in the Catalytic Chain by Serine Reduces Enzymatic Activity. *J. Biol. Chem.* **259**, 11180–11183.
- 38. Salemme, F. R. (1985), Engineering Aspects of Protein Structure. *Ann. NY Acad. Sci.* **439**, 97–106.
- 39. Shiroishi, T., Evans, G. A., Appella, E., and Ozato, K. (1985), In Vitro Mutagenesis of a Mouse MHC Class I Gene for the Examination of Structure–Function Relationships. *J. Immunol.* **134**, 623–629.
- 40. Shi, J. P., and Fersht, A. R. (1984), Fidelity of DNA Replication Under Conditions Used for Oligodeoxynucleotide-Directed Mutagenesis. *J. Mol. Biol.* 177, 269–278.
- 41. Taylor, H. C., Komoriya, A., and Chaiken, I. M. (1985), Crystallographic Structure of an Active, Sequence-Engineered Ribonuclease. *Proc. Natl. Acad. Sci. USA* 82, 6423–6426.
- 42. Tikhonenko, T. I. (1984), Prospects and Achievements of Genetic Engineering in Development of Antiviral Vaccines. *Mol. Biol. (Mosk)* **18**, 176–188.
- 43. Vane, J., and Cuatrecasas, P. (1984), Genetic Engineering and Pharmaceuticals [News]. *Nature* **312**, 303–305.
- 44. Villafranca, J. E., Howell, E. E., Voet, D. H., Strobel, M. S., Ogden, R. C., Abelson, J. N., and Kraut, J. (1983), Directed Mutagenesis of Dihydrofolate Reductase. *Science* 222, 782–788.
- 45. Waye, M. M., Verhoeyen, M. E., Jones, P. T., and Winter, G. (1985), EcoK Selection Vectors for Shotgun Cloning into M13 and Deletion Mutagenesis. *Nucleic Acids. Res.* **13**, 8561–8571.
- 46. Wang, A., Lu, S. D., and Mark, D. F. (1984), Site-Specific Mutagenesis of the Human Interleukin-2 Gene: Structure–Function Analysis of the Cysteine Residues. *Science* **224**, 1431–1433.
- 47. Wells, J. A., Vasser, M., and Powers, D. B. (1985), Cassette Mutagenesis: An Efficient Method for Generation of Multiple Mutations at Defined Sites. *Gene* **34**, 315–323.
- 48. Winter, G., Carter, P., Waye, M. M., Blow, D. M., Wilkinson, A. J., Shi, J. P., and Fersht, A. R. (1984), Genetic Dissection of Tyrosyl-tRNA Synthetase. *Biochem. Soc. Trans.* 12, 224–225.

49. Winter, G., Fersht, A. R., Wilkinson, A. J., Zoller, M., and Smith, M. (1982), Redesigning Enzyme Structure by Site-Directed Mutagenesis: Tyrosyl tRNA synthetase and ATP binding. *Nature* **229**, 756–758.

- 50. Wilkinson, A. J., Fersht, A. R., Blow, D. M., Carter, P., and Winter, G. (1984), A Large Increase in Enzyme-Substrate Affinity by Protein Engineering. *Nature* **307**, 187–188.
- 51. Wilkinson, A. J., Fersht, A. R., Blow, D. M., and Winter, G. (1983), Site-Directed Mutagenesis as a Probe of Enzyme Structure and Catalysis: Tyrosyl-tRNA Synthetase Cysteine-35 to Glycine-35 Mutation. *Biochemistry* **22**, 3581–3586.